

A Review On Anti-Thrombotic Agents Derived From Snake Venom Protein

Hibah A. Alharbi¹, Lujain M. Alabbas¹, Raghad I. Sumnudi¹, Shaima Felemban¹ and Yosra Alhindi²

¹Pharm D candidate, College of pharmacy, Umm Al-Qura University, Makkah, Saud Arabia.

²Department of Pharmacology and Toxicology, College of Medicine, Umm Al-Qura University, Makkah, Saudi Arabia

Abstract

Background

Snakes have always attracted scientists and it caused awe and fear because of the harmful strength of its toxins and the components of various toxins from one snake to another. In 2017, the World Health Organization (WHO) listed snake venom as a neglected high-priority disease, with snakes causing up to 2.7 million venomous bites, nearly 100,000 victims, and nearly three times the number of human disabilities. Snake venoms are mixtures of various protein components engaged in different functional activities on a variety of physiological target. In this review we will focus on, and the gather updated information about the antithrombotic effect of different snake venoms on their mechanism of action and therapeutic properties.

Objective

1. Snake venom background and therapeutics properties.
2. The effects of snake bite.
3. The mechanism of haemostasis.
4. Types of antithrombotic agents derived from snake venom proteins.

Conclusion

Snake venoms are a group of diverse compounds and because of their diversity, they lead to many effects that have different effects on the body, causing an imbalance and mastery of the victim. Just as these components have toxic effects, they have clinical and therapeutic effects. The previous research shown the clinical benefits of snake venom, such as the drug captopril used to treat high blood pressure, And the defibrase used for the prevention of thrombotic disease. Therefore, snake venom components need several studies. They are still unidentified due to the difficulty of obtaining them adequate.

Key words: Snake venoms, antithrombotic, haemostasis, venom proteins.

Introduction

Snake venom

Snakes have always been a source of attraction to scientists [1]. Moreover, it caused awe and fear not only because of their elegant movement of its limbs but also

because of the harmful strength of its toxins and the components of various toxins from one snake to another [2]. In 2017, the World Health Organization (WHO) listed snake venom as a neglected high-priority disease, with snakes causing up to 2.7 million venomous bites, nearly 100,000 victims,

and nearly three times the number of human disabilities [3,4]. Snake venoms are a huge mixture of different protein components, each protein in it acting on different functional activities on a set of varied physiological goals [5]. These venoms vary extensively between and within snake species [6]. Snakebite can cause many hazardous pathologies related to

neurotoxic, cytotoxic, and hemotoxic effects of the venom [6,7]. Both (Table.1 and Figure.1) demonstrates the most famous types of poisonous snakes responsible for the largest mortality in the world [8]. The diagnosis of poisoning is a pure clinical skill with no diagnostic tool kit available yet [9].

Table.1 The most poisonous snakes.

This table shows the most poisonous snakes (Family and Country) [10].

Snake	Common Name	Part in Figure1	Family	Country
Echis Ocellatus	West African Saw-Scaled Viper	B	Viperidae	West African
Bitis Arietans	Puff Adder	C	Viperidae	Savannah and Grasslands from Morocco, Western Arabia and Africa Except For The Sahara and Rainforest Regions
Naja Naja	Cobra	D	Elapidae	India, Pakistan, Bangladesh, Sri Lanka, Nepal, And Bhutan
Bungarus Caeruleus	Common Krait	E	Elapidae	Indian Subcontinent
Daboia Russelii	Western Russell's Viper	F	Viperidae	Asia (Common in India)
Bothrops Atrox	Common Lancehead	G	Viperidae	The Tropical Lowlands of North of the Americas East Of the Andes
Bothrops Asper	Terciopelo	H	Viperidae	Distribution from Southern Mexico To Northern South America.
Oxyuranus Scutellatus	Papuan Taipan	I	Elapidae	North & East Australia and New Guinea island.

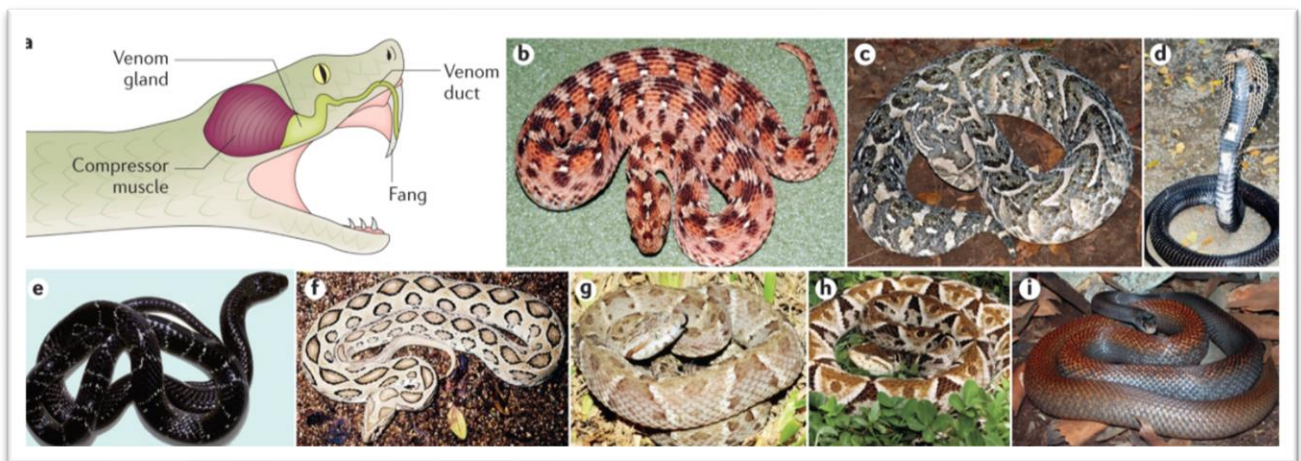


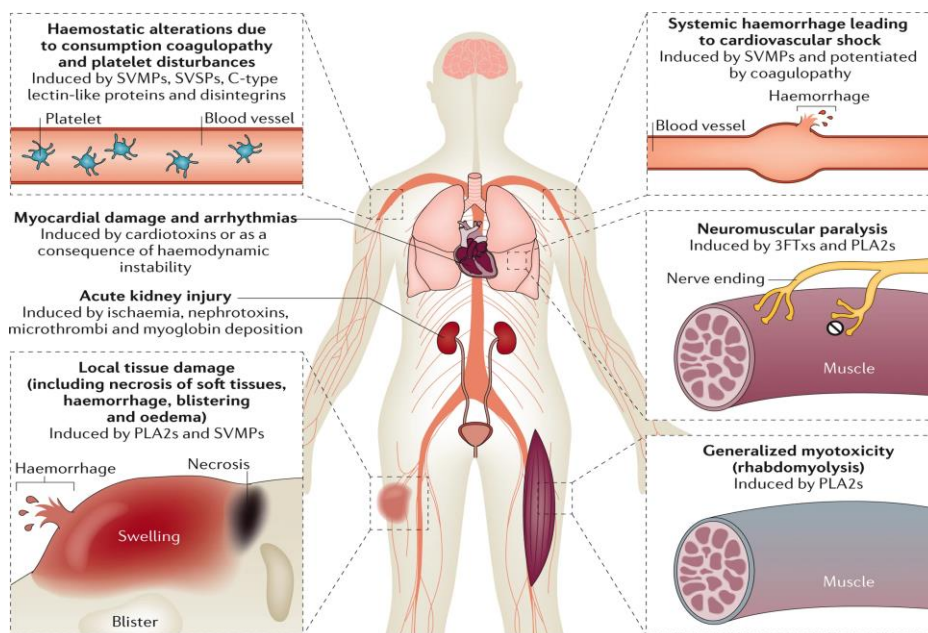
Figure.1 Venomous snakes.

This figure shows the Schematic illustration of the venom system in a snake of the family Viperidae and the most poisonous snakes [10].

The effects of snake bite

When snakes inject their venom, they use their tusks from the anterior part of maxillary bones (Figure.1) [11,12]. Depending on how big the canines are, the

toxin is injected under the skin or intramuscularly [13]. The toxins exert local effects on adjacent tissues once they are delivered, while others spread uniformly through the lymphatic system and blood vessels., allowing the toxins to act in several organs (Figure. 2) [14]. Therefore, we notice here that the spread of poison in the body causes poisoning in many organs, and every poisoning that occurs is caused by a component of snake venom [15].



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Figure.2 snake bite.

This figure shows the Action of snake venom toxins on different body systems [10,20]. Multiple changes can occur after a snake bite such as local swelling, local necrosis, non-specific early systemic symptoms (vomiting, headache, abdominal pain, critical diarrhoea, and fall with unrecordable blood pressure), spontaneous haemorrhage, neurotoxic effect (dizziness, headache, and fainting), myotoxic effect (myopathic tumours in skeletal muscle), cardiac effect (rapid pulse, low blood pressure and shock), renal failure [16,17,18,19,20].

Snake venom components

Snakes have multiple types of venom glands, which are nine species, including a family Elapidae and Viperidae [21]. Snake venom contains a hundred of different compounds [22]. These compounds are composed of organic, peptides, enzymes, proteins, chemicals, inorganic cations (sodium, zinc, calcium, magnesium, potassium), carbohydrates, free amino acids, and lipids [6,23]. The composition of the snake venom protein may change according to types of family and class, and this change affects the effectiveness of the protein and its strength [6]. Snake venom consists of four groups of proteins [20,24]. The first major protein group present in most types of venoms namely: Snake

venom metalloproteinases (SVMP), Three-finger toxins (3FTx), Phospholipases A2 (PLA2), and snake venom serine proteases (SVSP) [20]. The second group commonly exists but to a less amount than the first group: Kunitz peptides (KUN), L-amino acid oxidases (LAAO), Disintegrin (DIS), C-type lectins (CTL), Cysteine-Rich Secretory Proteins (CRiSP), and Natriuretic peptides (NP) [5,6,10]. The third group presents in little amount than the second group: acetylcholinesterase, vascular endothelial growth factor (VEGF), and venom nerve growth factor (VNGF) [5,6,10]. Last group is rare: Galactose-binding proteins, aminopeptidases or warping, and Cobra venom factors (CVF) [5,6,10]. Moreover, not all proteins present in all snakes venomous [5,6,10].

In our research, we will discuss the updated information about the antithrombotic effect of various snake toxins depending on their mechanism of action and therapeutic properties.

Method

A review was done by searching the databases as PubMed, google scholar and Medline. Key words included: "Snake venom", "Snake bite", "Venom anticoagulant" and "anticoagulant proteins. The updated search period is from 2016 to 2021.

Haemostasis and Thrombosis systemic

Haemostasis is the mechanism in our human body that includes several steps, and this results in what is called a "plug" that closes the damaged site in the blood vessels to protect against loss of blood [16]. Haemostasis can occur in two stages: primary and secondary [16]. Primary haemostasis is aimed to form a weak platelet plug, which is achieved by four steps [16]. Vasoconstriction is the first step occur after damaging of blood vessels and occur within thirty minutes

[16,25,26,27,28,29]. First, a vascular spasm of the blood vessels occurs, which stimulates vasoconstriction induced by endothelin-1 (a strong vasoconstrictor produced by the injured endothelium), injured endothelium revealed Sub-endothelial collagen, von Willebrand factor (vWF), and produces ATP and inflammatory mediators [16]. Platelet adhesion is the second step, it is a mechanism which platelets start rolling through the blood vessels and attach to the revealed subendothelial vWF and collagen in damaging areas, leading to platelet adherence and closing the injury [16,29]. In more specific, phospholipid bilayer of platelet membranes is rich with G protein receptors, especially Gp 1b-9 receptor that bind to vWF within the endothelium by making a link between them. [16]. Once linked, the platelet is activated by a set of processes [16]. Platelet activation is the third step, it happens when platelet binds to vWF, its activation passes by two processes [16]. The first is the shape of platelets will undergo an irreversible transformation from smooth discs to multi-pseudopodal, which dramatically expands the surface area [16]. The second is releasing of cytoplasmic granules of platelets [16]. There are two pathways of platelet activation that occur through thrombin [16]. One of them that thrombin stimulates platelets immediately by binding to a protease-activated receptor through proteolytic cleavage [16]. Another way is that thrombin releasing of cytoplasmic granules, including serotonin, thromboxane A2, platelet activation factor, and ADP [16,29]. The published ADP will link with P2Y1 and P2Y12 receptors on platelet membranes [16]. Binding to the P2Y1 receptor will P2Y1 receptor will aids to platelet aggregation by making conformational [16,29]. Binding to the P2Y12 receptor will aid to induce the coagulation pathway [16]. Moreover, the binding of ADP to its receptors on the

platelet membrane will induce Gp IIb/IIIa complex which is important for platelet-to-platelet aggregation and platelet-to-endothelial adherence [16]. Platelets produce TXA₂, which enhances vasoconstriction and platelet aggregation [16]. As a result, the platelet activation mechanism will make the zone eligible for platelet aggregation [16]. The last step of primary hemostasis is platelet aggregation [16]. When platelet activated the Gp IIb/IIIa receptors will bind to vWF and fibrinogen [16]. Then platelet will start binding to each other and will form a weak platelet plug [16,29]. Finally, primary haemostasis occurs to prevent bleeding temporarily until secondary haemostasis start [16]. Secondary haemostasis is defined as the transformation of fibrinogen into fibrin, which stabilizes the soft weak platelet plug and makes it a hard, insoluble fibrin clot [4]. This is done by three processes [16]: (1) activation of clotting factors, (2) conversion of prothrombin to thrombin, and (3) conversion of fibrinogen to fibrin. Within haemostasis, there are 3 coagulation pathways: intrinsic, extrinsic, and common pathways [16]. These processes are achieved by using one of the pathways; the extrinsic or the intrinsic pathway which focus on the activation of FX and then completes their processes by the common pathway [16]. In secondary haemostasis, calcium ions are required for all processes because it plays an important role in all 3 pathways [4]. The extrinsic pathway includes FVII and tissue factor (TF) [16]. It is initiated when TF binds to FVII, activating FVII to FVIIa, forming a TF-FVIIa complex [16]. This complex activates FX. TF-FVIIa complex can also activate FIX of the intrinsic pathway, which is called the alternate pathway [16]. Once FX is activated to FXa by the TF-FVIIa complex, the cascade continues the common pathway [16]. The intrinsic pathway includes the Hageman FXII, FXI, FIX, and FVIII [16]. The process is initiated

when FXII meets the exposed subendothelial collagen and becomes activated to FXIIa. FXIIa activates FXI to FXIa, and FXIa activates FIX to FIXa. FIXa works in combination with activated FVIIIa to activate FX [16]. Once FX is activated by the FIXa-FVIIIa complex, the cascade continues the common pathway [16]. The common pathway is initiated by the activation of FXa [16]. FXa combines with FVa and calcium on phospholipid surfaces to create a prothrombinase complex activating prothrombin (FII) into thrombin (FIIa) [16]. This activation of thrombin occurs by serine protease cleaving of prothrombin [16]. Now, thrombin activates FXIIIa [16]. FXIIIa crosslinks with fibrin forming the stabilized clot [16]. Both intrinsic and extrinsic pathways are shared to continue coagulation with the common pathway [4].

Thrombosis is the formation of clots in the blood vessels and leads to partial or complete obstruction, and a decrease in the amount of blood flowing in the blood vessels, because of changes in persistent blood components [4,16]. Thrombosis occurs more frequently in the veins but is also present in the arteries, capillaries, and heart [19,21,50]. The thrombosis occurred in; 1, change in the diameter of the vessel lumen with following variations in the blood current; a/ dilation of the vascular lumen: varicose veins, aneurysms, or Nodar periarteritis.; b/ contraction of the vascular lumen: ligation, compression of the vessel from the outside, or obstruction inside the vessel.; 2, certain metabolic alterations like cachexia, obesity, experimental hypothyroidism, hyperparathyroidism.; 3, the action of poisons may cause disintegration of blood elements including polymorphonuclear leucocytes.; 4, change in the physicochemical balance of the blood proteins like tissue necrosis, hyperallergic, infectious diseases.; 5, alterations in the thickness of the plasma: cholera, leukemia,

hyperglobulia, chlorosis.; 6, change of the line between blood and tissue due to lesions of the vascular wall, phlebitis, arteriosclerosis; supposedly in certain cases of irritation of nerves and in postoperative shock, in particular, when acting on the veins of the abdominal and genital region.;

7, changes resulting from the action of excess heat, coldness, or radiation [2]. And in the following figure (Figure.3) shows the pathways of coagulation and the mechanism of different components of anticoagulant snake venoms [30].

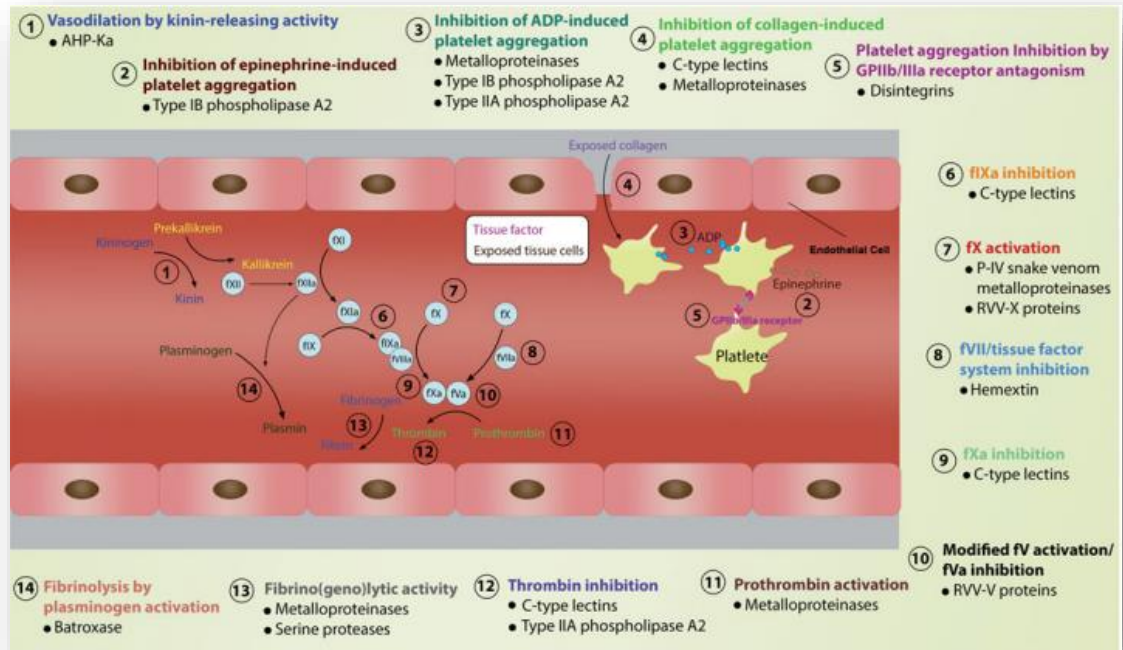


Figure.3 Anticoagulant Mechanism of Snake Venom on coagulation pathway.

This figure shows the pathways of coagulation and the mechanism of different components of anticoagulant snake venoms [30].

Enzymatic Anticoagulant Proteins Phospholipase A2 (PLA2)

Phospholipase A2 is a superfamily it is a toxic enzyme in snake venom [30]. It is a small protein and has worked in two bonds: The first bond is sn-2 fatty acyl bond is binding to sn-2 fatty acyl of phospholipids membrane to make lysophospholipids and free fatty acids [30]. The second bond is the sn-2 ester bond PLA2 acts to hydrolyze the

sn-2 ester bond of glycerophospholipids is a structural lipid bilayer cellular membrane [31, 32,33]. They main divided for PLA2 into two groups: Old World snakes (group I) and New World snakes (group II) they are different in disulfide bond pattern both two groups have comparable molecular weight (13–15 kDa) [18, 34]. Studies have established the enzyme Phospholipase A2 has an anticoagulant effect by two mechanisms: non-enzymatic and enzymatic [35]. Enzymatic mechanism it is inhibiting the formation of prothrombinase complex (FVa, Ca²⁺, phospholipids, and FXa) [18]. The non-enzymatic mechanism it is inhibiting the formation of thrombin by inhibiting the FXa (Figure.4) [18,36].

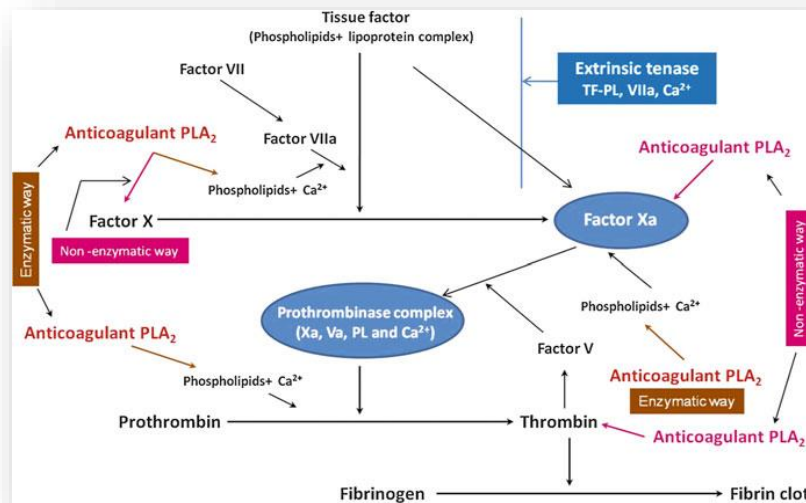


Figure.4 Anticoagulant Mechanism of Phospholipase A2.

This figure shows two mechanisms for Phospholipase A2 as anticoagulant: Non enzymatic and enzymatic [18].

Snake-venom metalloproteases (SVMP)

Snake Venom Metalloproteinases (SVMPs) are a group of zinc-dependent enzymes described in the venoms of snakes [35,36,37,38,39, 40]. They have several biological activities such as proteolytic degradation of fibrinogen and fibrin, inhibition of platelet aggregation and

induce haemorrhage [35]. Snake venom metalloproteinases have a significant antithrombotic and haemorrhagic activities in vivo [40]. They are divided into three groups based on domain structure and molecular weight (Table.2) [40]. These compounds work to inhibit the platelet aggregation by blocking collagen binding to integrin a2b1 on platelets [40].

Table.2 The Summary of Metalloproteinases groups.

This table shows the Metalloproteinases groups based on domain structure and molecular weight and their Haemorrhagic effect [40].

Groups	Molecular weight	Domain structure	Hemorrhagic effect
P-I	20–30 kDa	Only a proteinase domain	Weak
P-II	30–60 kDa	Proteinase and disintegrin domains	High
P-III	60–100 kDa	Cysteine-rich domain, proteinase domain and disintegrin-like domain	Higher than other groups

Serine proteases (SVSP)

SVSP is a group of enzymes derived from snake venom, and like other

components of snake venom, it performs several functions such as platelet aggregation [37], blood coagulation [7,26,30,37,41], fibrinolysis [7,26,30,41],

hypotensive [26,37], neurotoxicity [26], and anticoagulant effect [26]. These enzymes are divided into two groups [26]. The first group, which is predominant, is thrombin-like enzymes, and it stimulates the blood clotting process and forms fibrin by cleaving the A α and/or B β chains of fibrinogen [7,26,30,41]. The other group of SVSP is protein C activators, this activated protein C inactivates FV/FVa and FVIII/FVIIIa and releases a tissue-type plasminogen activator. It also interacts with plasminogen activator inhibitors to make the fibrinolysis effect [26].

L-amino acid oxidases (LAAO)

L-amino acid oxidases (LAAOs) are flavoproteins found in Elapidae and Viperidae, but especially in Crotaline [11,32,42]. These enzymes represent a variety of biological activities in the victim such as cytotoxicity, myotoxicity, and edema, which induce clinical symptoms of envenomation [11, 23,40,43]. L-amino acid oxidases are shown a broad range of pharmacological activities such as antimicrobial, platelet aggregation (anticoagulant), cellular apoptosis

(anticancer), and anti-HIV activity [1,44]. L-amino acid oxidases (LAAOs) which activates the stereospecific de-lamination of the L-amino acid layer into alpha-ketogenic acid, which produces ammonia and hydrogen peroxide (Figure.5) [32,33,40,45,46]. These enzymes make up one to nine percent of total venom proteins [32,36,40]. Some studies have described LAAOs with inhibiting effects on platelets, while others have reported LAAOs with stimulating effects [36,40]. platelet aggregation induced by H₂O₂ production and Consequent synthesis of thromboxane A₂ [13,36,40]. While the inhibitory mechanism of LAAO induced by four methods [23]. 1/Reduced attachment of ADP in platelets exposed to H₂O₂ [40].; 2/Interposition of peroxide at the site of interaction between activated platelet integratrin GP IIb/IIIa and fibrinogen [40].; 3/Collagen, Thrombin, ADP, and Arachidonic Acid [40].; 4/Selectively suppresses FIX activity [36]. And some study told they have both anti-aggregation and de-aggregation effects on the rabbit and human platelets [40].

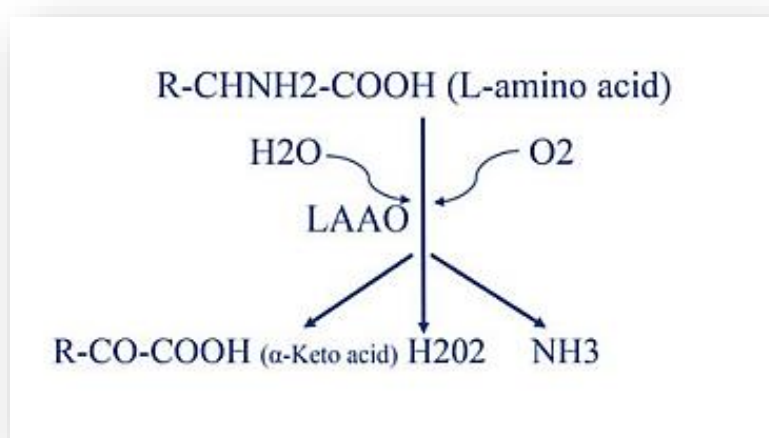


Figure.5 The Mechanism of L-Amino Acid Oxidases Enzymes.

This figure shows the de-amination mechanism of L-amino acid coat to an

alpha-keto acid, that produce ammonia and hydrogen peroxide [23].

Nucleotidases

They are hydrolytic enzymes have high molecular mass ranging from 73 to 100 kDa and involved in the cleavage of nucleic acid derivatives and substrates [6]. They are present in many types of snake venoms but only few of them were isolated due to their limited concentrations in venoms [6]. Their primary function is to release adenosine by hydrolyse 5'-nucleotides to nucleosides which exhibit antiplatelet and hypotensive effect [6]. Also, adenosine contributes to biodistribution of toxins, immobilization, increase vascular permeability, inhibit neurotransmitter release, that leads to sedation, bradycardia, hypotension, and locomotor depression [6]. These changes can enhance synergistically the anticoagulant action of certain toxins as ADPases, phospholipases A2 and Disintegrins [6]. Another study found that Nucleotidases show antiplatelet effect by interact with blood FIX in coagulation cascade [37,47].

Non-Enzymatic Anticoagulant Proteins

Three-finger toxins (3FTx)

Three-finger toxins of snake venom (3FTxs) are a significant family of non-

enzymatic proteins toxins commonly found in elapids venom [3,22,27,31,37,41,48]. The name "Three-Finger toxin" comes from its special structure, consisting of 57 to 82 amino acid remainders and folded into three crooked rings extending from a compressed, hydrophobic core [27, 37,41,48]. The core is clamped by four preserved disulphide bridges forming a three-finger shape (Figure.6) [27, 37,41,48]. The functional multiplicity of 3FTxs through diversity in amino acid sequences and other structural changes, that leads to bind to different receptors and exhibit a wide variety of biological effects such as neurotoxic, cytotoxic, cardiotoxic effects, and hypotensive effect [3,22,27,31,37,41,48]. 3-FTxs is one of the biggest family of snake venom toxins, more than 800 sequences of 3FTxs that have been hoarded due to the diversity in amino acid sequences and other structural changes [3,27]. One of its many interesting functions is antiplatelet and anticoagulation [27,48]. Where more than one toxin substance extracted from 3ftx has been reported, it has an anti-platelet or anticoagulant effect [27,48]. In this table (table.3) we can see the names of the toxic substances, their Origin, and their mechanism of action [27,48,49,50,51,52].

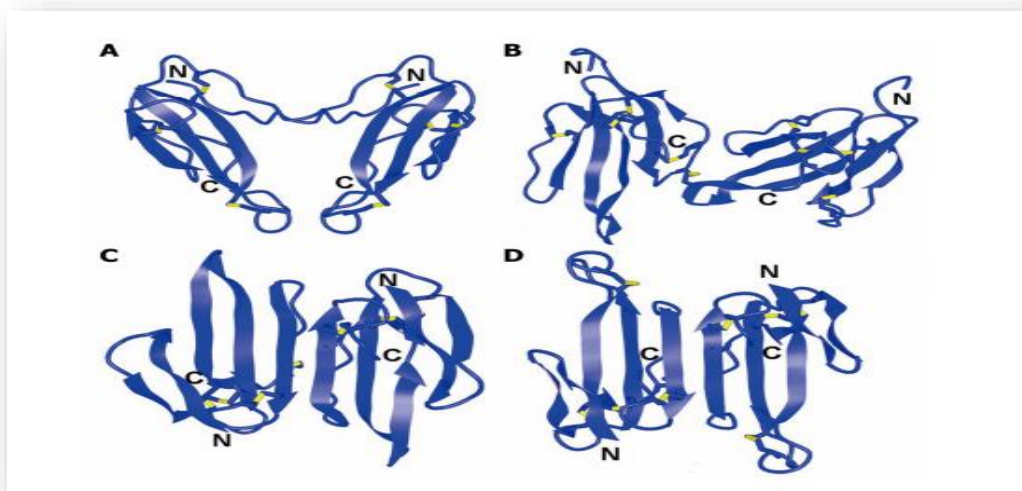


Figure.6 The Structure of Three-Finger Toxin.

This figure shows the variation of structure lead to variety of biological effects; (A) Homodimer of α -cobratoxin. (B) Irditoxin. (C) 0Haditoxin. (D) κ -bungarotoxin. (N) &(C) N- and C-terminus, yellow color expresses bisulfide bonds [48].

Table.3 Antiplatelet and Anticoagulant of Three-Finger Toxins.

This table shows the names of Antiplatelet and Anticoagulant of Three-Finger Toxins, their origin, and their Mechanism of action [27,48].

3FTx	Origin	Mechanism of Action
Anti-Platelet		
Dendroaspis	Dendroaspis Jamesoni Kaimosae	Inhibits ADP-mediated platelet aggregation and inhibits binding of the purified alpha-IIb/beta-3 platelet fibrinogen receptor (ITGA2B/ITGB3) to paralyzed fibrinogen.
S5C1		
Thrombostatin	Dendroaspis Angusticeps	
γ -bungarotoxin	Bungarus Multicinctus	
TA-bm16		
NTL2		
KT-6.9	Naja Kaouthia	<ul style="list-style-type: none"> • Inhibit inductive ADP, thrombin, and arachidonic acid. • Hinder platelet aggregation by adenosine diphosphate. • Inhibit platelet induced through the P2Y12 receptor.
Anti-Coagulant		
Hemextin AB complex	Hemachatus Haemachatus	<ul style="list-style-type: none"> • Links to FVIIa only • Inhibition of ETC formation by Prevent interaction between FVII and TF
Exactin		
Ringhalexin	Naja Atra	<ul style="list-style-type: none"> • Inhibition of ETC formation. • Links to FVIIa and TF complex that interfere with the FX link.
Najalexin		
Ophiolexin	Ophiophagus Hannah	

Kunitz peptides (KUN)

Kunitz-Type Proteinase Inhibitors was the first protease inhibitor was discovered in snake venoms 1972 by Takahashi and colleagues, it contains 60 amino acid, 6 cysteine residues and stabilize by 3 disulphide bridges [6,18]. KUN was singled out from the venom of *D. russelli* and reported to exist in Elapidae and Viperidae snake venoms [6]. This family of protease inhibitors has structural similarity to bovine pancreatic trypsin inhibitor BPTI, which is responsible for serine proteases inhibition such as activated FX, elastase, trypsin, thrombin, and chymotrypsin [6,9]. It has a wide range of biological activities like ion channels blockage especially voltage-gated potassium channels which may lead to anticoagulation, fibrinolysis, and inflammation [6, 53].

C-type lectins (CTL)

C-type lectins are non-enzymatic proteins and Ca²⁺ dependent carbohydrate proteins [42-49]. These family of proteins are found in many different tissue types and are primarily involved in adherence and immunity and signalling related to inflammation [18]. All these family proteins portion homology in their carbohydrate-recognition domains (CRDs) [18,49], Though, all these family has C-type lectins and high difference in amino acid sequence so-called C-type lectin fold [18]. Platelets are an integral part of the haemostat [18]. The first step of the constitutes blood coagulation cascade is Platelet aggregation [18]. C-type lectins bind to multiple integrins and receptor to each block or inhibit platelet aggregation: von Willebrand Factor, Thrombin, GPIb (receptor glycoprotein Ib), FIX and/or FX [14,17,18,34]. C-type lectins binding to von Willebrand Factor to form complex to inhibit/prevent activation of platelet aggregation [14,34]. Another mechanism is binding to receptor GPIb (is platelet

membrane glycoprotein Ib) and inhibits the formation of thrombosis [14,18]. Binding with FX and/or FIX formation complex to act as anticoagulants [34].

Disintegrins (DIS)

Disintegrins are a family of small cysteine-rich peptides, was discovered first in 1987 and found to inhibit the platelet aggregation by various agonists, including collagen, ADP, sodium arachidonate, and epinephrine that neither affected the shape change nor the cyclic adenosine monophosphate (cAMP) level [4,12]. Disintegrins bind to integrins through several methods: 1/Arginine-glycine-aspartate (RGD)-containing peptides resulting in an active site that modulates the integrin activity [12,40].; 2/It inhibit ADP-induced platelet aggregation [40].; 3/Acts as α IIb β 3 antagonists because it is playing a big role in platelet functions, haemostasis, and arterial thrombosis [24,28].

Conclusion

Snake venoms are a group of diverse compounds and because of their diversity, they lead to many effects that have different effects on the body, causing an imbalance and mastery of the victim [6]. On the other hand, snake venom is also considered to have clinical benefits that have been shown by previous research, as there are currently many drugs that have been extracted from snake venom, such as the drug captopril used around the world, which inhibits the angiotensin-converting enzyme (ACE) to treat high blood pressure [37], And also the defibrase is a product based on batroxobin which is a procoagulant SVSP that extracted from snake venom of *Bothrops atrox* and used for the prevention of thrombotic disease [30]

Just as these components have toxic effects, they have clinical and therapeutic effects [6]. Therefore, snake venom components need several studies [6]. They are still unidentified due to the difficulty of

obtaining them adequately [6]. In this research we gathered information about the effect of snake venom on the blood. However, two classes of venom proteins, snakecs and disintegrins have been shown to specifically target receptors expressed on platelets, endothelial cells, phagocytes, tumor cells, thus affecting cell-matrices and cell-cell interactions. the potential applications of venom field of integrin-related diseases, especially arterial thrombosis, angiogenesis, tumor progression and septic inflammation. Moreover, novel components are being developed as a safer antithrombotic agent with minimal side effects, such as thrombocytopenia and bleeding.

List of abbreviations

3FTx- Three-finger toxins.

a2b1- Analysis of Integrin Alpha2Beta1.

ADP- Adenosine diphosphate.

ATP- Adenosine 5-TriPhosphate.

A α - Fibrinogen Alpha Chain (Gene).

B β - Fibrinogen Beta Chain (Gene).

cAMP- Cyclic Adenosine Monophosphate.

CRiSP- Cysteine-Rich Secretory Proteins.

CTL- C-type lectins.

CVF- Cobra venom factors.

DIS- Disintegrin.

ECT- Extrinsic Tenase Complex.

FII / FIIa- Factor two (Prothrombin) / Activated Factor two (thrombin).

FIX / FIXa- Factor nine (Christmas) / Activated Factor nine.

FV / FVa- Factor Five (Labile) / Activated Factor Five.

FVII / FVIIa- Factor Seven (Stable) / Activated Factor Seven.

FVIII / FVIIIa- Factor eight (Antihemophilic) / Activated Factor eight.

FX / FXa- Factor ten (Stuart) / Activated Factor ten.

FXI / FXIa- Factor eleven (Plasma Thromboplastin Antecedent) / Activated Factor eleven.

FXII / FXIIa- Factor twelve (Hageman) / Activated Factor twelve.

FXIII / FXIIIa- Factor Thirteen / Activated Factor Thirteen.

H₂O₂- Hydrogen Peroxide.

KDa- Killodaitons.

KUN- Kunitz peptides.

LAAO- L-amino acid oxidases.

NP- Natriuretic peptides.

PLA₂- Phospholipases A₂.

RGD- Arginine-glycine-aspartate.

SVMP- Snake venom metalloproteinases.

SVSP- snake venom serine proteases.

TF- Tissue Factor.

TXA₂- Thromboxane A₂.

VEGF- vascular endothelial growth factor.

VNGF- venom nerve growth factor.

Vwf- Von Willebrand Factor.

WHO- World Health Organization.

References

1. Abdelkafi-Koubaa, Z., Aissa, I., Morjen, M., Kharrat, N., El Ayeb, M., Gargouri, Y., ... & Marrakchi, N. (2016). Interaction of a snake venom L-amino acid oxidase with different cell types membrane. *International journal of biological macromolecules*, 82, 757-764.

2. Ashorobi, D., & Fernandez, R. (2019). Thrombosis. StatPearls [Internet].
3. Babenko, V. V., Ziganshin, R. H., Weise, C., Dyachenko, I., Shaykhutdinova, E., Murashev, A. N., ... & Utkin, Y. (2020). Novel Bradykinin-Potentiating Peptides and Three-Finger Toxins from Viper Venom: Combined NGS Venom Gland Transcriptomics and Quantitative Venom Proteomics of the Azemiops feae Viper. *Biomedicines*, 8(8), 249.
4. Barmore, W., Bajwa, T., & Burns, B. (2020). Biochemistry, clotting factors. StatPearls [Internet].
5. Bocian, A., & Hus, K. K. (2020). Antibacterial properties of snake venom components. *Chemical Papers*, 74(2), 407-419.
6. Boldrini-Franca, J., Cologna, C. T., Pucca, M. B., Bordon, K. D. C. F., Amorim, F. G., Anjolette, F. A. P., ... & Arantes, E. C. (2017). Minor snake venom proteins: Structure, function and potential applications. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1861(4), 824-838.
7. Carone, S. E., Menaldo, D. L., Sartim, M. A., Bernardes, C. P., Caetano, R. C., da Silva, R. R., ... & Sampaio, S. V. (2018). BJSPP, a novel serine protease from Bothrops jararaca snake venom that degrades fibrinogen without forming fibrin clots. *Toxicology and applied pharmacology*, 357, 50-61.
8. Casewell, N. R., Jackson, T. N., Laustsen, A. H., & Sunagar, K. (2020). Causes and consequences of snake venom variation. *Trends in pharmacological sciences*.
9. Chapeaurouge, A., Silva, A., Carvalho, P., McCleary, R. J., Modahl, C. M., Perales, J., ... & Mackessy, S. P. (2018). Proteomic deep mining the venom of the red-headed krait, *Bungarus flaviceps*. *Toxins*, 10(9), 373.
10. Charvat, R. A., Strobel, R. M., Pasternak, M. A., Klass, S. M., & Rheubert, J. L. (2018). Analysis of snake venom composition and antimicrobial activity. *Toxicon: official journal of the International Society on Toxinology*, 150, 151-167. <https://doi.org/10.1016/j.toxicon.2018.05.016>.
11. Costal-Oliveira, F., Stransky, S., Guerra-Duarte, C., de Souza, D. L. N., Vivas-Ruiz, D. E., Yarlequé, A., ... & Braga, V. M. (2019). L-amino acid oxidase from *Bothrops atrox* snake venom triggers autophagy, apoptosis and necrosis in normal human keratinocytes. *Scientific reports*, 9(1), 1-14.
12. David, V., Succar, B. B., De Moraes, J. A., Saldanha-Gama, R. F. G., Barja-Fidalgo, C., & Zingali, R. B. (2018). Recombinant and chimeric disintegrins in preclinical research. *Toxins*, 10(8), 321.
13. de Queiroz, M. R., de Sousa, B. B., da Cunha Pereira, D. F., Mamede, C. C. N., Matias, M. S., de Moraes, N. C. G., ... & de Oliveira, F. (2017). The role of platelets in hemostasis and the effects of snake venom toxins on platelet function. *Toxicon*, 133, 33-47.
14. Drabeck, D. H., Rucavado, A., Hingst-Zaher, E., Cruz, Y. P., Dean, A. M., & Jansa, S. A. (2020). Resistance of South American opossums to vWF-binding venom C-type lectins. *Toxicon*, 178, 92-99.
15. Estevão-Costa, M. I., Sanz-Soler, R., Johanningmeier, B., & Eble, J. A. (2018). Snake venom components in medicine: From the symbolic rod of Asclepius to tangible medical research

- and application. *The international journal of biochemistry & cell biology*, 104, 94-113.
16. Garmo, C., & Burns, B. (2018). Physiology, clotting mechanism.
 17. Glatz, J. C., & Luiken, J. F. (2018). Dynamic role of the transmembrane glycoprotein CD36 (SR-B2) in cellular fatty acid uptake and utilization. *Journal of lipid research*, 59(7), 1084-1093.
 18. Paniagua, D., Vergara, I., Boyer, L., Alagón, A., Gopalakrishnakone, P., Inagaki, H., ... & Vogel, C. W. (2017). Role of lymphatic system on snake venom absorption. In *Snake Venoms* (pp. 453-474). Springer Dordrecht.
 19. Gupta, N., Zhao, Y. Y., & Evans, C. E. (2019). The stimulation of thrombosis by hypoxia. *Thrombosis research*, 181, 77-83.
 20. Gutiérrez, J. M., Calvete, J. J., Habib, A. G., Harrison, R. A., Williams, D. J., & Warrell, D. A. (2017). Snakebite envenoming. *Nature reviews Disease primers*, 3(1), 1-21.
 21. Hardy, T. J., & Bevis, P. M. (2019, February 1). *Deep vein thrombosis. Surgery (United Kingdom)*. Elsevier Ltd. <https://doi.org/10.1016/j.mpsur.2018.12.002>.
 22. Hegde, R. P., Rajagopalan, N., Doley, R., & Kini, R. M. (2016). 13 Snake Venom Three-Finger Toxins. *Handbook of Venoms and Toxins of Reptiles*, 287.
 23. Hiu, J. J., & Yap, M. K. K. (2020). Cytotoxicity of snake venom enzymatic toxins: phospholipase A2 and l-amino acid oxidase. *Biochemical Society Transactions*, 48(2), 719-731.
 24. Huang, J., Li, X., Shi, X., Zhu, M., Wang, J., Huang, S., ... & Jin, J. (2019). Platelet integrin $\alpha\text{IIb}\beta\text{3}$: signal transduction, regulation, and its therapeutic targeting. *Journal of hematology & oncology*, 12(1), 1-22.
 25. Jesudasan, J. E., & Abhilash, K. P. P. (2019). Venomous snakebites: Management and anti-snake venom. *Current Medical Issues*, 17(3), 66.
 26. Gogoi, D., Arora, N., Kalita, B., Sarma, R., Islam, T., Ghosh, S. S., ... & Mukherjee, A. K. (2018). Anticoagulant mechanism, pharmacological activity, and assessment of preclinical safety of a novel fibrin (ogen)olytic serine protease from leaves of *Leucas indica*. *Scientific reports*, 8(1), 1-17.
 27. Kini, R. M., & Koh, C. Y. (2020). Snake venom three-finger toxins and their potential in drug development targeting cardiovascular diseases. *Biochemical Pharmacology*, 114, 105.
 28. Kuo, Y. J., Chung, C. H., Pan, T. Y., Chuang, W. J., & Huang, T. F. (2020). A Novel $\alpha\text{IIb}\beta\text{3}$ Antagonist from Snake Venom Prevents Thrombosis without Causing Bleeding. *Toxins*, 12(1), 11.
 29. LaPelusa, A., & Dave, H. D. (2019). Physiology, hemostasis.
 30. Latinović, Z., Leonardi, A., Koh, C. Y., Kini, R. M., Trampuš Bakija, A., Pungercar, J., & Križaj, I. (2020). The procoagulant snake venom serine protease potentially having a dual, blood coagulation Factor V and X-activating activity. *Toxins*, 12(6), 358.
 31. Modahl, C. M. (2016). Evolution and Biological Roles of Three-Finger Toxins in Snake Venoms.
 32. Moga, M. A., Dimienescu, O. G., Arvătescu, C. A., Ifteni, P., & Pleș, L.

- (2018). Anticancer activity of toxins from bee and snake venom—an overview on ovarian cancer. *Molecules*, 23(3), 692.
33. Molla, G., Melis, R., & Pollegioni, L. (2017). Breaking the mirror: L-amino acid deaminase, a novel stereoselective biocatalyst. *Biotechnology advances*, 35(6), 657-668.
34. Ning, W., Yuanyuan, L., Lipeng, Z., Xiang, L., & Chunhong, H. (2020). Targeted identification of C-type lectins in snake venom by 2DE and Western blot. *Toxicon*, 185, 57-63.
35. Olaoba, O. T., Dos Santos, P. K., Selistre-de-Araujo, H. S., & de Souza, D. H. F. (2020). Snake Venom Metalloproteinases (SVMPs): A structure-function update. *Toxicon*: X, 100052.
36. Paloschi, M. V., Pontes, A. S., Soares, A. M., & Zuliani, J. P. (2018). An update on potential molecular mechanisms underlying the actions of snake venom L-amino acid oxidases (LAAOs). *Current medicinal chemistry*, 25(21), 2520-2530.
37. Péterfi, O., Boda, F., Szabó, Z., Ferencz, E., & Bába, L. (2019). Hypotensive snake venom components—A mini-review. *Molecules*, 24(15), 2778.
38. Post, Y., Puschhof, J., Beumer, J., Kerckamp, H. M., de Bakker, M. A., Slagboom, J., ... & Kazandjian, T. D. (2020). Snake venom gland organoids. *Cell*, 180(2), 233-247.
39. Randolph, C. E., Blanksby, S. J., & McLuckey, S. A. (2019). Toward complete structure elucidation of glycerophospholipids in the gas phase through charge inversion ion/ion chemistry. *Analytical chemistry*, 92(1), 1219-1227.
40. Rashidi, R., Valokola, M. G., Rad, S. Z. K., Etemad, L., & Roohbakhsh, A. (2018). Antiplatelet properties of snake venoms: a mini review. *Toxin Reviews*.
41. Rey-Suárez, P., Saldarriaga-Córdoba, M., Torres, U., Marin-Villa, M., Lomonte, B., & Núñez, V. (2019). Novel three-finger toxins from *Micrurus dumerilii* and *Micrurus mipartitus* coral snake venoms: Phylogenetic relationships and characterization of Clarkitoxin-I-Mdum. *Toxicon*, 170, 85-93.
42. Rincon-Filho, S., Naves-de-Souza, D. L., Lopes-de-Souza, L., Silvano-de-Oliveira, J., Ferreyra, C. B., Costal-Oliveira, F., ... & Chávez-Olórtegui, C. (2020). *Micrurus surinamensis* Peruvian snake venom: Cytotoxic activity and purification of a C-type lectin protein (Ms-CTL) highly toxic to cardiomyoblast-derived H9c2 cells. *International Journal of Biological Macromolecules*, 164, 1908-1915.
43. Sanhajariya, S., Duffull, S. B., & Isbister, G. K. (2018). Pharmacokinetics of snake venom. *Toxins*, 10(2), 73.
44. Slagboom, J., Kool, J., Harrison, R. A., & Casewell, N. R. (2017). Haemotoxic snake venoms: their functional activity, impact on snakebite victims and pharmaceutical promise. *British journal of haematology*, 177(6), 947-959.
45. Tasoulis, T., & Isbister, G. K. (2017). A review and database of snake venom proteomes. *Toxins*, 9(9), 290.
46. Ullah, A. (2020). Structure–Function Studies and Mechanism of Action of Snake Venom L-Amino Acid Oxidases. *Frontiers in pharmacology*, 11, 110.

47. Ullah, A., Masood, R., Ali, I., Ullah, K., Ali, H., Akbar, H., & Betzel, C. (2018). Thrombin-like enzymes from snake venom: Structural characterization and mechanism of action. *International journal of biological macromolecules*, 114, 788-811.
48. Utkin, Y. N. (2019). Last decade update for three-finger toxins: Newly emerging structures and biological activities. *World journal of biological chemistry*, 10(1), 17.
49. Varki, A., Cummings, R. D., Esko, J. D., Stanley, P., Hart, G. W., Aebi, M., ... & Seeberger, P. H. (2017). *Study Guide. In Essentials of Glycobiology [Internet]. 3rd edition. Cold Spring Harbor Laboratory Press.*
50. Vazhappilly, C. G., Ansari, S. A., Al-Jaleeli, R., Al-Azawi, A. M., Ramadan, W. S., Menon, V., ... & Radhakrishnan, R. (2019). Role of flavonoids in thrombotic, cardiovascular, and inflammatory diseases. *Inflammopharmacology*, 27(5), 863-869.
51. Williams, P. E., Klein, D. R., Greer, S. M., & Brodbelt, J. S. (2017). Pinpointing double bond and sn-positions in glycerophospholipids via hybrid 193 nm ultraviolet photodissociation (UVPD) mass spectrometry. *Journal of the American Chemical Society*, 139(44), 15681-15690.
52. Xie, C., Albulescu, L. O., Still, K., Slagboom, J., Zhao, Y., Jiang, Z., ... & Kool, J. (2020). Varespladib inhibits the phospholipase A2 and coagulopathic activities of venom components from hemotoxic snakes. *Biomedicines*, 8(6), 165.
53. Youngman, N. J., Walker, A., Naude, A., Coster, K., Sundman, E., & Fry, B. G. (2020). Varespladib (LY315920) neutralises phospholipase A2 mediated prothrombinase-inhibition induced by Bitis snake venoms. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 236, 108818.